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(54) Method for the extraction and purification of DNA.

(57) Method for the extraction and purification of DNA, starting from lambda phages, M13 phagemids, plasmids, cosmids or other like elements, which provides for three steps:

- charging the sample, which undergoes an exchange treatment, or lysis, followed by macrofiltration,
- the macrofiltrate is treated with a precipitant and the resulting solution is subjected to ultrafiltration,
- the DNA fixed to the filtration support of ultrafiltration is eluted and collected.

Lambda DNA, single-stranded DNA from phagemids, plasmidic DNA and cosmidic DNA obtained by the above method.

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## METHOD FOR THE EXTRACTION AND PURIFICATION OF DNA

This invention concerns a method to extract and purify DNA. To be more exact, the invention provides a method which can be reproduced automatically with modest execution times and with simple automation problems.

The invention can be employed to obtain DNA from lambda phages or the like, M13 phagemids and the like, and also from plasmids and cosmids and the like.

The journal *Nucleic Acids Research*, vol.15, number 23, 1987, includes on page 10047 an article headed, "A simple and fast method for preparing single-stranded DNA template suitable for sequencing".

The same journal in vol.16, number 7, 1988 includes on page 2873 another article headed, "A new and fast method for preparing high lambda DNA suitable for sequencing".

These articles teach the obtaining of DNA from phagemids and lambda phages respectively.

The methods disclosed in the above articles provide for at least one centrifuging step, and it is the inclusion of these centrifuging steps besides other factors which makes the automation of the method hard and inconvenient.

The above proposed methods arrange also to exchange the detergent CTAB (cetyl-trimethyl-ammonium bromide) with the sodium ion by means of re-suspension of the CTAB-DNA pellet, obtained after centrifuging, in a 1.2 M NaCl solution with the addition of two volumes of ethanol and further centrifuging.

Although these teachings have improved the state of the art by eliminating the separation of steps inherent in deproteinization by means of phenol, yet they entail the shortcoming of not enabling the process of obtainment to be automated simply, for the centrifuging in fact hinders or, in any event, makes very complex the automation of the process except with excessive costs and unjustified execution times.

To obviate these shortcomings and achieve further advantages which will become clear in the following description, such as the speed of the process and the ability to reproduce the results, as well as the yield, the present applicant has studied, tested and obtained the present method.

The method according to the invention is set forth in the main claim, whereas the dependant claims describe variants of the idea of the main solution.

The invention provides for the use of appropriate, differentiated filtration cycles instead of centrifuging.

The method according to the invention enables

plasmid DNA, cosmid DNA, phagemid DNA or lambda phage DNA to be obtained.

The DNA obtained with the method can be employed in further analyses of:

- 1) enzymatic restriction
- 2) cloning
- 3) enzymatic synthesis reactions (RNA polymerase and DNA polymerase)
- 4) sequencing reactions.

The yield of DNA by percentage, as compared to the methods described in the aforesaid publications making use of centrifuging, ranges from 80% to 100%.

According to the invention differentiated filtrations with differentiated functions are employed to obtain the required DNA.

The method according to the invention comprises at least the following steps:

step 1: charging the sample, which undergoes a first exchange treatment followed by macro-filtration.

According to the invention the exchange treatment can be differentiated in relation to the typology of the starting sample;

step 2: the macrofiltrate is treated with a precipitant and the resulting solution is subjected to ultrafiltration;

step 3: after washing, the DNA fixed to the filtration support is eluted.

Let us examine these individual steps one by one.

### Step 1.

After the charging, an anionic exchange resin (solution A), DEAE resin for instance, in a quantity of a volume equal to that of the sample, is added to the sample of lysate of phages or phagemids. Another analogous substance can be used instead of the anionic exchange resin.

The whole is re-mixed for a time long enough for the desired exchange to take place. This time may be 10 minutes at the ambient temperature, for instance.

For the bacterial culture, in order to obtain plasmidic DNA or cosmidic DNA the solution A which is added will contain an alkaline solution of lysis (ph  $\geq$  11) that comprises sodium desoxycolate (DOC) ( $\geq$  0.05%) or another like product.

After the necessary re-mixing for proper homogenization of the solution thus produced (mixing lasting for 2 or 3 minutes, for instance) at the ambient temperature a solution (solution B) containing an acid, acetic acid for instance, is added to

the bacterial culture alone so as to bring the pH to about 5 ( $\pm 2$ ).

The solution thus produced is mixed again for enough time to obtain formation of a macrofloc-  
 5 culate (complex of DOC/genomic DNA/cellular debris).

The invention provides for the use of desox-  
 ycolate (DOC) as a detergent for bacterial lysis,  
 instead of dodecylsulphate (SDS) as taught in the  
 state of the art when the sample contains plasmid  
 or cosmid, for the exchange treatment.

The essential advantage of the use of deter-  
 gent DOC in the process is the more efficient  
 precipitation of the detergent/ genomic DNA/cellular  
 10 debris complex with a slightly acid pH (pka of  
 DOC = 6.9 whereas the pka of SDS = 1-2).

Precipitation of the detergent in an acid am-  
 bient ( $< 6.9$ ) enables the detergent and that which  
 is associated with it to be wholly removed, as  
 opposed to what happens when SDS is employed.

In fact, with the detergent SDS used in the  
 most common processes to obtain plasmidic DNA  
 or cosmidic DNA it is necessary, besides acidifica-  
 tion, to increase the concentration of sodium ions  
 and to lower the temperature.

If SDS is used to proceed thereafter to the step  
 of precipitation with CTAB, as described in the  
 cited state of the art, it is necessary to dilute the  
 filtrate by three times beforehand.

Moreover, even when the filtrate is diluted by  
 three times, the detergent SDS, if employed to  
 produce the solution, is not wholly eliminated, and  
 this causes a successive inefficient formation of the  
 DNA-CTAB complex.

With macrofiltration through a macroporous  
 diaphragm the DEAE resin is separated out with all  
 the anionic impurities associated therewith in the  
 case of a lysate of phages/phagemids and the  
 macrofloculate is separated out in the case of a  
 bacterial culture (plasmids or cosmids).

In fact, depending on the starting element, the  
 impurities will have been adsorbed by the ex-  
 change resin DEAE or will be the object of macro-  
 precipitate generated by the DOC detergent of  
 lysis.

The macroporous diaphragm may consist, for  
 instance, of compressed gauze or an equivalent  
 filter means.

In the case of lysate of phages or phagemids  
 the macroporous diaphragm may be supported ad-  
 vantageously by a nitrocellulose filter, for instance,  
 of an 0.2 micron type or the like (Teflon,  
 borosilicates, etc.).

## Step 2.

To the filtrate is added the precipitant solution

(solution C) containing, for instance, cetyl-trimethyl-  
 ammonium bromide (CTAB), which is a cationic  
 detergent of a low ionic force that forms a micellar  
 complex with the polyanion DNA.

This precipitant solution enables the  
 DNA/CTAB micellar complex to be formed after  
 brief mixing for homogenization.

Where plasmids or cosmids are used as the  
 starting material, the macrofiltrate can be neutral-  
 10 ized beforehand and be treated with an enzyme,  
 RNasi A, for instance, to digest the RNA contained  
 in the lysate before addition of the precipitant.

The solution thus obtained and containing the  
 micellar complex then undergoes ultrafiltration.

The ultrafiltration can be carried out with any  
 known microporous support; for instance, a micro-  
 porous support consisting of borosilicate glass  
 (DURAN type or the like with a porosity of 4) has  
 been usefully employed and has enabled the DNA-  
 CTAB micellar complex to be retained satisfac-  
 15 torily.

The second filtration, being of an ultrafiltration  
 and inert type, makes possible:

1) adhesion of the DNA-cationic detergent  
 (CTAB for instance) micellar precipitate;

2) washing and exchange of the detergent  
 with the ions, Na for instance,

3) recovery of the DNA in the form of salt,  
 for instance sodium salt, in an aqueous solution.

## Step 3.

The DNA (lambda, phagemid, plasmid, cosmid)  
 35 being now immobilized on the filter (as a result of  
 Step 2), it is now eluted.

However, this elution takes place after a series  
 of washings of the filter, as follows for instance:

- a first wash is carried out in an aqueous solution  
 of a low ionic force to eliminate excess detergent;

- a second wash is carried out with a mixture of  
 40 alcohol and ionic solution, for instance of a NaCl  
 0.2 M/70% ethanol type, to exchange the DNA-  
 CTAB into DNA-Na;

- a third wash is then carried out with a mixture of  
 45 alcohol and a solution of a low ionic force to  
 remove the excess salts, for instance of a NaCl  
 type (for example, 70% ethanol, 30% water).

After having dried every trace of alcohols care-  
 50 fully from the filter, the DNA is then recovered by  
 washing the filter with an aqueous solution of a low  
 ionic force (water for instance).

The DNA thus obtained, whether lambda DNA,  
 single-stranded DNA from phagemids or plasmidic  
 DNA or cosmidic DNA, can now be used for subse-  
 55 quent purposes.

## Claims

- 1 - Method for the extraction and purification of DNA, starting from lambda phages, M13 phagemids, plasmids, cosmids or other like elements, characterized in that it provides for three steps: 5
- charging the sample, which undergoes an exchange treatment, or lysis, followed by macrofiltration, 10
  - the macrofiltrate is treated with a precipitant and the resulting solution is subjected to ultrafiltration,
  - the DNA fixed to the filtration support of ultrafiltration is eluted and collected. 15
- 2 - Method as claimed in Claim 1, in which in the case of lambda phages and M13 phagemids the exchange treatment is carried out with an anionic exchange resin, DEAE for instance. 15
- 3 - Method as claimed in Claim 1, in which in the case of plasmids or cosmids an alkaline solution of lysis ( $\text{pH} \geq 11$ ) is added which contains a detergent, for instance sodium desoxycolate (DOC). 20
- 4 - Method as claimed in Claim 1 or 3, in which a solution containing acid is added to bring the pH to about 5 ( $\pm 2$ ) so as to obtain a macrofloculate (for instance, DOC/genomic DNA/cellular debris). 25
- 5 - Method as claimed in any claim hereinbefore, in which the macrofiltration takes place with a macroporous diaphragm. 30
- 6 - Method as claimed in any of Claims 1, 2 or 5, in which in the case of lambda phages or M13 phagemids the macroporous diaphragm is supported by a nitrocellulose filter of 0.2 microns or the like. 35
- 7 - Method as claimed in any of Claims 1, 3, 4 or 5, in which the macrofiltrate derived from plasmids or cosmids is neutralized at least to reduce the acidity. 40
- 8 - Method as claimed in any claim hereinbefore, in which the macrofiltrate is treated with an enzyme suitable to digest RNA. 40
- 9 - Method as claimed in any claim hereinbefore, in which a precipitant solution containing a cationic detergent, for instance CTAB (cetyltrimethyl-ammonium bromide), is added to the macrofiltrate to obtain a micellar complex. 45
- 10 - Method as claimed in any claim hereinbefore, in which the micellar complex undergoes ultrafiltration, successive washing and elution of the retained portion with recovery of the DNA. 50
- 11 - Lambda DNA, single-stranded DNA from phagemids, plasmidic DNA and cosmidic DNA obtained according to one or another of the features of the claims hereinbefore. 55



DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl.5)
A,D	NUCLEIC ACIDS RESEARCH, vol. 16, no. 7, 11th April 1988, pages 2873-2884, IRL Press Ltd, Oxford, GB; G. MANFIOLETTI et al.: "A new and fast method for preparing high quality lambda DNA suitable for sequencing" * Materials and method" *	1	C 12 N 15/10 C 12 P 19/34
A	EP-A-0 240 191 (SEIKO INSTRUMENT INC.) * Page 1, column 1, line 35 - column 2, line 27 *	1	
A,D	NUCLEIC ACIDS RESEARCH, vol. 15, no. 23, 1987, page 10047, IRL Press Ltd, Oxford, GB; G. DEL SAL et al.: "A simple and fast method for preparing single stranded DNA template suitable for sequencing"		
			TECHNICAL FIELDS SEARCHED (Int. Cl.5)
			C 12 N C 12 P
The present search report has been drawn up for all claims			
Place of search THE HAGUE		Date of completion of the search 23-03-1990	Examiner VAN PUTTEN A.J.
<b>CATEGORY OF CITED DOCUMENTS</b> X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons & : member of the same patent family, corresponding document			